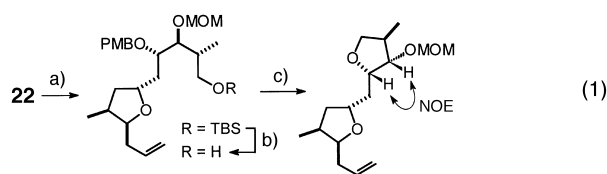


As mentioned above, the other members of the amphidinolide T family, that is, amphidinolide T1–T3, differ from T4 (**1**) mainly or exclusively in the C-12 to C-14 region of the macrocycle;^[3] because the stereocenter at C-13 in **30** is appropriately set, however, the synthesis of these compounds is just a matter of a different timing of the final deprotection steps and/or simple inversion reactions. Therefore it is expected that the synthesis summarized above will give access to these valuable marine natural products as well. This and related aspects are subject to ongoing studies in our laboratory.

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This assignment was ultimately confirmed by the conversion of **22** into **1** and comparison of the data of the synthetic sample with those of the natural product.

- [1] a) M. Ishibashi, J. Kobayashi, *Heterocycles* **1997**, *44*, 543–572; b) T. K. Chakraborty, S. Das, *Curr. Med. Chem. Anti Cancer Agents* **2001**, *1*, 131–149.
- [2] Specifically, amphidinolides H and N exhibit antitumor potency similar to that of the spongistatins, see ref. [1] and references therein.
- [3] J. Kobayashi, T. Kubota, T. Endo, M. Tsuda, *J. Org. Chem.* **2001**, *66*, 134–142.
- [4] For total syntheses of other amphidinolides see: a) D. R. Williams, W. S. Kissel, *J. Am. Chem. Soc.* **1998**, *120*, 11198–11199; b) D. R. Williams, B. J. Myers, L. Mi, *Org. Lett.* **2000**, *2*, 945–948; c) D. R. Williams, K. G. Meyer, *J. Am. Chem. Soc.* **2001**, *123*, 765–766; d) H. W. Lam, G. Pattenden, *Angew. Chem.* **2002**, *114*, 526–529; *Angew. Chem. Int. Ed.* **2002**, *41*, 508–511; e) T. K. Chakraborty, S. Das, *Tetrahedron Lett.* **2001**, *42*, 3387–3390; f) R. E. Maleczka, L. R. Terrell, F. Geng, J. S. Ward, *Org. Lett.* **2002**, *4*, 2841–2844; g) B. M. Trost, J. D. Chisholm, S. T. Wroblewski, M. Jung, *J. Am. Chem. Soc.* **2002**, *124*, 12420–12421.
- [5] a) M. Tsuda, T. Endo, J. Kobayashi, *J. Org. Chem.* **2000**, *65*, 1349–1352; b) T. Kubota, T. Endo, M. Tsuda, M. Shiro, J. Kobayashi, *Tetrahedron* **2001**, *57*, 6175–6179.
- [6] a) D. S. Brown, M. Bruno, R. J. Davenport, S. V. Ley, *Tetrahedron* **1989**, *45*, 4293–4308; b) A. Schmitt, H.-U. Reißig, *Eur. J. Org. Chem.* **2001**, 1169–1174.
- [7] a) E. Negishi, V. Bagheri, S. Chatterjee, F.-T. Luo, J. A. Miller, A. T. Stoll, *Tetrahedron Lett.* **1983**, *24*, 5181–5184; b) E. Negishi, F. Liu, in *Metal-catalyzed Cross-coupling Reactions* (Eds.: F. Diederich, P. J. Stang), Wiley-VCH, Weinheim, **1998**, pp. 1–47; c) Y. Tamaru, H. Ochiai, F. Sanda, Z.-I. Yoshida, *Tetrahedron Lett.* **1985**, *26*, 5529–5532; d) P. Knochel, R. D. Singer, *Chem. Rev.* **1993**, *93*, 2117–2188.
- [8] a) T. M. Trnka, R. H. Grubbs, *Acc. Chem. Res.* **2001**, *34*, 18–29; b) A. Fürstner, *Angew. Chem.* **2000**, *112*, 3140–3172; *Angew. Chem. Int. Ed.* **2000**, *39*, 3012–3043; c) M. Schuster, S. Blechert, *Angew. Chem.* **1997**, *109*, 2124–2144; *Angew. Chem. Int. Ed. Engl.* **1997**, *36*, 2037–2056.
- [9] U. S. Racherla, Y. Liao, H. C. Brown, *J. Org. Chem.* **1992**, *57*, 6614–6617.
- [10] J. R. Gage, D. A. Evans, *Org. Synth.* **1990**, *68*, 83–91.
- [11] For a similar sequence see: K. Makino, K. Kimura, N. Nakajima, S. Hashimoto, O. Yonemitsu, *Tetrahedron Lett.* **1996**, *37*, 9073–9076.
- [12] a) R. Noyori, *Asymmetric Catalysis in Organic Synthesis*, Wiley, New York, **1994**; b) T. Ohkuma, R. Noyori in *Comprehensive Asymmetric Catalysis, Vol. 1* (Eds.: E. N. Jacobsen, A. Pfaltz, H. Yamamoto), Springer, Berlin, **1999**, pp. 199–246.
- [13] D. A. Evans, M. D. Ennis, D. J. Mathre, *J. Am. Chem. Soc.* **1982**, *104*, 1737–1739.
- [14] The corresponding TMS enol ether is labile and leads to erratic results.
- [15] For a review on the stereoselective synthesis of natural products with polysubstituted THF units see: U. Koert, *Synthesis* **1995**, 115–132.
- [16] Protection of the alcohol **22** as a PMB ether [PMB = *para*-methoxybenzyl; a) PMBBBr, KHMDS], cleavage of the terminal TBS group [b) TBAF, THF] and treatment of the resulting alcohol with c) I₂, PPh₃, and imidazole delivers the tetrahydrofuran shown in Equation (1) rather than the expected primary iodide, compare O. R. Martin, F. Yang, F. Xie, *Tetrahedron Lett.* **1995**, *36*, 47–50. Its relative stereochemistry was elucidated by NOE measurements, from which the stereochemistry of **22** formed with L-Selectride can be deduced.
- [17] For another advanced example see: A. Fürstner, H. Weintritt, *J. Am. Chem. Soc.* **1998**, *120*, 2817–2825.
- [18] These expectations stem from results in the following projects: a) A. Fürstner, K. Radkowski, C. Wirtz, R. Goddard, C. W. Lehmann, R. Mynott, *J. Am. Chem. Soc.* **2002**, *124*, 7061–7069; b) A. Fürstner, F. Jeanjean, P. Razon, *Angew. Chem.* **2002**, *114*, 2203–2206; *Angew. Chem. Int. Ed.* **2002**, *41*, 2097–2101; c) A. Fürstner, T. Dierkes, O. R. Thiel, G. Blanda, *Chem. Eur. J.* **2001**, *7*, 5284–5296; d) A. Fürstner, O. R. Thiel, N. Kindler, B. Bartkowska, *J. Org. Chem.* **2000**, *65*, 7990–7995; e) A. Fürstner, T. Müller, *J. Am. Chem. Soc.* **1999**, *121*, 7814–7821; f) A. Fürstner, K. Langemann, *J. Am. Chem. Soc.* **1997**, *119*, 9130–9136; g) A. Fürstner, K. Langemann, *J. Org. Chem.* **1996**, *61*, 3942–3943.
- [19] a) M. Scholl, T. M. Trnka, J. P. Morgan, R. H. Grubbs, *Tetrahedron Lett.* **1999**, *40*, 2247–2250; b) J. Huang, E. D. Stevens, S. P. Nolan, J. L. Pedersen, *J. Am. Chem. Soc.* **1999**, *121*, 2674–2678; c) L. Ackermann, A. Fürstner, T. Weskamp, F. J. Kohl, W. A. Herrmann, *Tetrahedron Lett.* **1999**, *40*, 4787–4790; d) A. Fürstner, L. Ackermann, B. Gabor, R. Goddard, C. W. Lehmann, R. Mynott, F. Stelzer, O. R. Thiel, *Chem. Eur. J.* **2001**, *7*, 3236–3253.
- [20] K. Takai, Y. Hotta, K. Oshima, H. Nozaki, *Tetrahedron Lett.* **1978**, 2417–2420; b) T. Okazoe, J. Hibino, K. Takai, H. Nozaki, *Tetrahedron Lett.* **1985**, *26*, 5581–5584; c) for a highly successful application in a case where the Wittig reaction had also failed see: A. Fürstner, O. Guth, A. Düffels, G. Seidel, M. Liebl, B. Gabor, R. Mynott, *Chem. Eur. J.* **2001**, *7*, 4811–4820.
- [21] S. Matsubara, M. Sugihara, K. Utimoto, *Synlett* **1998**, 313–315.
- [22] R. K. Boeckman, Jr., P. Shao, J. J. Mullins, *Org. Synth.* **2000**, *77*, 141–152.

Tuning PCR Specificity by Chemically Modified Primer Probes**

Michael Strerath and Andreas Marx*

Dedicated to Professor Peter Welzel
on the occasion of his 65th birthday

Since the publication of the first draft of the human genome sequence in 2001, the discovery of genomic dissimilarities such as single nucleotide polymorphisms (SNPs) between different individuals has been a main focus of many research

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Supporting information for this article is available on the WWW under <http://www.angewandte.org> or from the author.

efforts.^[1] This is an specially intriguing issue since it is becoming increasingly apparent that variable effects of drugs on different patients or predisposition for various diseases can often be linked to variations in the genome sequence.^[1] In the future, the exact knowledge of medicinally relevant nucleotide variations may allow any therapy to be adapted to the genetic equipment of the individual and would abolish treatment with drugs that are ineffective or cause severe side effects.^[1] Methods that allow time- and cost-efficient verification of nucleotide variations should lead to further advances in pharmacogenetics.^[1,2]

Many methods for the detection of nucleotide variations in genes have been described to date.^[2] Each method exhibits advantages and disadvantages, and thus no methodology has prevailed so far. Most known methods are applied after amplification of the target genome sequence through the polymerase chain reaction (PCR). Thus, after isolation of the genetic material, the sequence of interest has to be amplified by PCR prior to the analytical step determining nucleotide variation.^[2] Methods for the direct detection of nucleotide variations through PCR are rare. Allele-specific amplification (ASA) determines nucleotide variations through either the presence or absence of a DNA product obtained through PCR amplification.^[3] The principle of ASA is based on the formation of matched or mismatched primer template complexes through allele-specific primer probes. From matched 3'-primer termini PCR amplification by a DNA polymerase proceeds, while a mismatch should obviate amplification. Nevertheless, there have been many reports indicating the low selectivity of this approach, thus necessitating further tedious time- and cost-intensive optimizations.^[4] Therefore, any means of increasing the selectivity of allele-specific PCR amplification should have significant impact on the reliability and robustness of direct SNP analysis by PCR.

Recently, we reported that nucleotide insertion selectivity of a DNA polymerase can be increased by chemical means by using 4'-alkylated nucleotides.^[5] The supplemental small modifications are believed to cause less flexibility within the DNA polymerase substrate complex and thus decrease the tolerance for formation of geometrically altered nascent nucleotide pairs.^[5,6] Based on this model, we speculated that the flexibility at the 3'-primer end can be decreased as well through these modifications, resulting in lower DNA polymerase propensity for extension of geometrically altered mismatched primer template complexes. Such a system could in turn be the basis for the development of a novel highly allele-specific PCR approach. Here we show that this concept is indeed applicable and report on a significant increase of the specificity of PCR by the use of chemically modified primer probes. Our results represent a major advancement in the development of robust methods for the detection of nucleotide variation through PCR.

We first synthesized DNA strands bearing 4'-modified thymidine residues at the 3'-terminal position and subsequently used these DNA strands as primers in primer extension assays.^[5b,7] First, we screened several thermostable DNA polymerases in their ability to extend from 4'-modified primer ends, which are either bound to the template according to the Watson–Crick rule or which cause a single mismatch

through a mutated template nucleotide (Figure 1).^[8] To mimic a cycle in PCR we employed reaction conditions commonly used in standard PCR protocols and used reaction buffers as recommended by the supplier.^[7] Screening several commer-

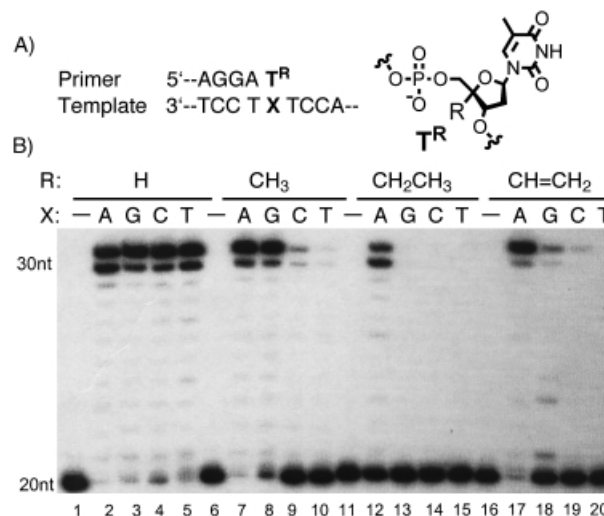


Figure 1. Effects of 4'-modifications on the primer extension catalyzed by Vent⁺. A) Section of the primer template sequences employed. B) Results of primer extension reactions obtained when canonical (X = A) or noncanonical (X = G, C, or T) primer ends were employed. The respective template nucleobases and 4'-substituents are depicted in the figure. Lanes 1, 6, 11, 16: primer only.^[7]

cially available thermostable DNA polymerases we identified Vent⁺ DNA polymerase (exonuclease-deficient mutant, Vent⁺) as the best mismatch discriminating enzyme in primer extensions employing 4'-modified oligonucleotides. The data obtained for Vent⁺ employing unmodified probes reveal that the enzyme significantly promotes primer extension even from mismatched primers under the applied conditions (Figure 1, lanes 2–5).

However, 4'-modifications at the primer termini cause in most incidences significantly increased discrimination between matched and mismatched primer template complexes (Figure 1, lanes 6–20). Discriminative primer extension turns out to be dependent on the bulk of the 4'-modification. Thus, 4'-methylated primer strands are extended efficiently by the DNA polymerase in the case of primer template T^{Me}/A as well as T^{Me}/G, while extension from T^{Me}/C and T^{Me}/T is significantly hampered (Figure 1, lanes 6–10). Increasing the bulk of the 4'-modification by employment of ethyl modifications led to significantly increased apparent mismatch discrimination but nevertheless, at the cost of efficiency for extension of matched complexes (Figure 1, lanes 11–15). Interestingly, 4'-vinyl-modified primers exhibit good discrimination under the conditions applied, combined with a significantly increased extension propensity in the matched case compared to the 4'-ethylated probes (Figure 1, lanes 16–20). These combined features render the 4'-vinyl modification the most interesting for further investigations.

For genotyping needs we next envisaged the synthesis of the corresponding probes bearing 4'-vinylated 2'-deoxycytidine moieties. Furthermore, subsequent investigations of their interplay with DNA polymerases should reveal whether the

observed effects on mismatch discrimination are exclusively linked to thymidine residues. Following a published procedure for the synthesis of **1**,^[9] we synthesized the modified solid support **2** bearing adequately protected 4'-vinylated 2'-deoxycytidine (Figure 2 A).^[10]

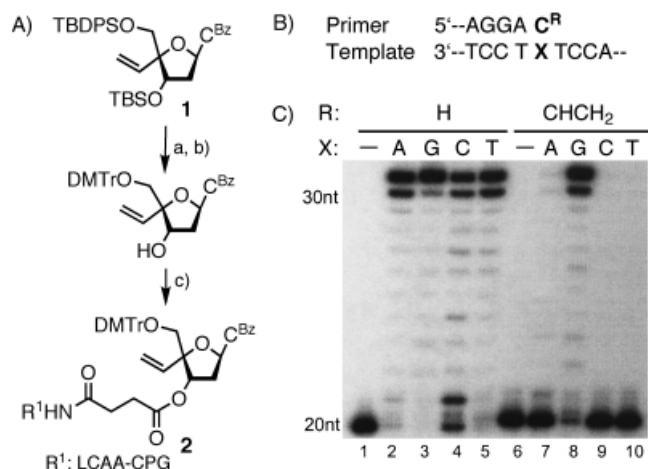


Figure 2. Synthesis of probes containing 4'-vinylated 2'-deoxycytidine and their effects on primer extension. A) Synthesis of solid supports bearing 4'-modified 2'-deoxycytidines. Reagents and conditions: a) TBAF, THF, 25°C, 4 h; b) DMTrCl, cat. DMAP, pyridine, 25°C, 7 h, 55% (two steps); c) succinylated LCAA-CPG, DMAP, EDC, NEt₃, pyridine, 25°C, 16 h; then 4-nitrophenol, 25°C, 14 h, then piperidine, 25°C, 5 min, then acetic anhydride, DMAP, pyridine, 25°C, 14 h. C^{Bz} = benzoyl-protected cytosine, TBS = tert-butyldimethylsilyl; TBDPS = tert-butyldiphenylsilyl, DMTr = 4,4'-dimethoxytrityl; DMAP = 4-dimethylaminopyridine; LCAA-CPG = long-chain alkyl amine modified controlled pore glass; EDC = 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride. B) Section of the primer template sequences employed. C) Results of primer extension reactions obtained when canonical (X = G) or noncanonical (X = A, C, or T, respectively) primer ends were employed. The respective template nucleobases and 4'-substituents are depicted in the figure. Lanes 1 and 6: primer only.^[7]

The required 4'-modified primer probes were synthesized by means of automated DNA synthesis by employing the modified solid supports. Subsequent primer extension studies with 4'-vinylated primer and Vent⁻ conducted under identical conditions as above led to results similar to those obtained with 4'-modified thymidines. The 4'-vinylated 2'-deoxycytidine residues likewise show significant beneficial effects on the apparent mismatch discrimination by enzymatic primer extension.

Encouraged by these promising findings we next explored whether the results of the primer extension can be further developed to a highly allele-specific PCR. First, we studied PCR amplification by Vent⁻ of a 63nt gene fragment of the human acid ceramidase, which contains nucleotide variations at a single position opposite the 3'-terminus of the respective primer probe.^[8] To explore the impact of 4'-vinyl modifications in primer probes on the discrimination of ASA we studied every possible single nucleotide variation in the template strand. As shown in Figure 3, usage of unmodified primer probes resulted in little discrimination between matched and single mismatched primer template complexes. Strikingly, employment of primer probes bearing 4'-vinylated thymidine residues resulted in a markedly diminished ampli-

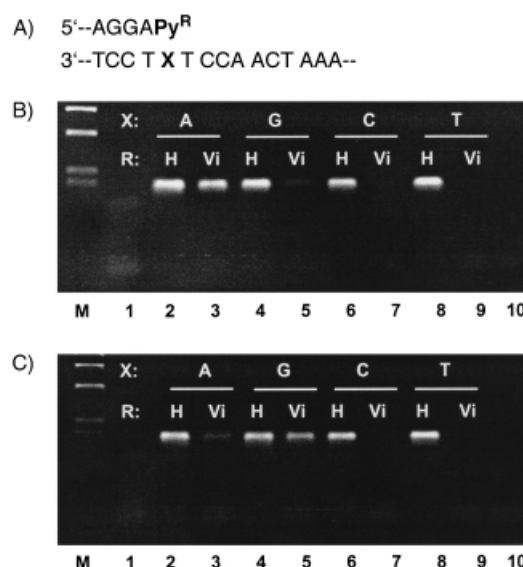


Figure 3. Effects of modified primer probes on the selectivity of PCR amplification. A) Section of sequences employed. Py^R = pyrimidine residues bearing 4'-hydrogen atom or 4'-vinyl modifications; X = respective template nucleobase as depicted in B) and C). B) Results obtained with primer probes bearing 4'-vinylated thymidine residues. C) Results obtained with primer probes bearing 4'-vinylated 2'-deoxycytidine residues. M = size marker; Lanes 1, 10: Experiments conducted in the absence of template and in each case with unmodified primers or 4'-modified primers.^[7]

fication of DNA when the primer probes were mismatched. These effects are significant in the presence of primer pyrimidine/template purine mismatches (Tⁱ/G; Cⁱ/A), but nevertheless even more pronounced when pyrimidine/pyrimidine mismatched probes were present. In the latter cases the extent of product amplification was beyond the detection limit under the employed conditions (Figure 3). Noteworthy, it has been shown that discrimination through PCR of pyrimidine/purine mismatches at the 3'-primer end (e.g. T/G) is difficult to achieve; thus, our findings of significantly increased selectivity due to 4'-modified primer probes are a significant advancement in this respect.^[4]

To gain detailed quantitative insight into this process we employed radioactively labeled primer probes and analyzed the ratio of product amplification after every cycle.^[7] The obtained results clearly show the highly beneficial effects of our modified primer probes on PCR amplification selectivity (Figure 4). For instance, 4'-vinylated primer termini are proficiently elongated in mismatched cases with somewhat lower efficiency than the unmodified probes. Nevertheless, single mismatches at the primer ends prevent efficient amplification significantly more pronounced in the 4'-modified cases resulting in a more selective PCR amplification with respect to a single nucleotide variation at the primer end.

In summary, we report on effects of 4'-modified primer probes on single mismatch discrimination in allele-specific PCR amplification. We found that in all investigated single mismatches, significantly higher amplification selectivity is observed by use of 4'-vinylated primer probes and Vent⁻ DNA polymerase compared to cases where unmodified primers were used. This study reveals that small chemical modifications of DNA can trigger distinct enzymatic respons-

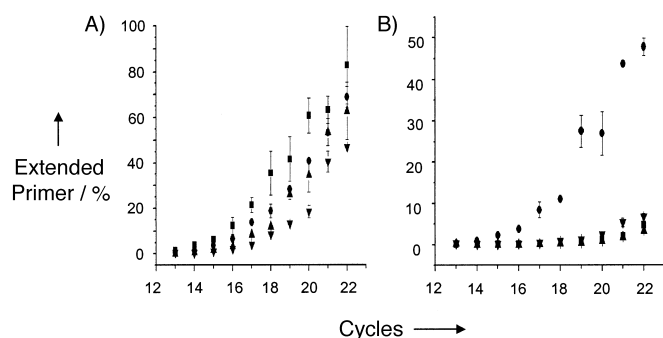


Figure 4. Quantification of PCR products. Results were obtained by employing the same conditions as shown in Figure 3 using 5'-³²P-labeled primer probes. Aliquots were removed after every cycle and subsequently analyzed by denaturing PAGE using phosphorimaging technology. The percentage of product relative to the overall radioactivity in one lane was plotted against the number of cycles. Error bars derive from repeated experiments. A) Results obtained with probes bearing unmodified thymidine residues at the 3'-primer end. B) Results obtained with primer probes bearing 4'-vinylated thymidine residues at the 3'-primer end. The respective primer/template duplexes were indicated by the following symbols: ● T^R/A, ▼ T^R/G, ▲ T^R/C, ■ T^R/T.^[7]

es that are not easily accessible with unmodified substrates. Considering recent developments in the use of real-time quantitative PCR as an analytical tool for detecting variations in nucleotides,^[11] these findings should have significant impact on the development of reliable and robust methods.

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- [1] For example: a) A. D. Ross, *Nature* **2000**, *405*, 857–865; b) J. J. McCarthy, R. Hilfiker, *Nat. Biotechnol.* **2000**, *18*, 505–508; c) W. E. Evans, M. V. Relling, *Science* **1999**, *286*, 487–491; d) P. W. Kley, E. S. Vesell, *Science* **1998**, *281*, 1820–1821.
- [2] For example: a) M. M. Shi, *Clin. Chem.* **2001**, *47*, 164–172; b) I. G. Gut, *Hum. Mutat.* **2001**, *17*, 475–492; c) M. Chicurel, *Nature* **2001**, *412*, 580–582; d) P. Y. Kwok, *Annu. Rev. Genomics Hum. Genet.* **2001**, *2*, 235–258.
- [3] D. Y. Wu, L. Ugozzoli, B. K. Pal, R. B. Wallace, *Proc. Natl. Acad. Sci. USA* **1989**, *86*, 2757–2760.
- [4] For example: a) Z. Guo, Q. Liu, L. M. Smith, *Nat. Biotechnol.* **1997**, *15*, 331–335; b) Y. Ishikawa, K. Tokunaga, K. Kashiwase, T. Akaza, K. Tadokoro, T. Juji, *Hum. Immunol.* **1995**, *42*, 315–318; c) P. R. Wenham, C. R. Newton, W. H. Price, *Clin. Chem.* **1991**, *37*, 241–244; d) C. R. Newton, A. Graham, L. E. Heptinstall, S. J. Powell, C. Summers, N. Kalsheker, J. C. Smith, A. F. Markham, *Nucleic Acids Res.* **1989**, *17*, 2503–2516.
- [5] a) D. Summerer, A. Marx, *Angew. Chem.* **2001**, *113*, 3806–3808; *Angew. Chem. Int. Ed.* **2001**, *40*, 3693–3695; b) D. Summerer, A. Marx, *J. Am. Chem. Soc.* **2002**, *124*, 910–911; c) M. Strerath, D. Summerer, A. Marx, *ChemBioChem* **2002**, *3*, 578–580.
- [6] E. T. Kool, *Annu. Rev. Biochem.* **2002**, *71*, 191–219.
- [7] Detailed experimental procedures as well as DNA sequences applied are provided in the Supporting Information.
- [8] All DNA polymerases experiments conducted were done in the sequence context of human acid ceramidase comprising the recently discovered mutation at A107. See: J. Bär, T. Linke, K. Ferlinz, U. Neumann, E. H. Schuchman, K. Sandhoff, *Hum. Mutat.* **2001**, *17*, 199–209.
- [9] N. Nomura, S. Shuto, M. Tanaka, T. Sasaki, S. Mori, S. Shigata, A. Matsuda, *J. Med. Chem.* **1999**, *42*, 2901–2908.
- [10] R. T. Pon in *Current Protocols in Nucleic Acids Chemistry* (Eds.: S. L. Beaucage, D. E. Bergstrom, G. D. Glick, R. A. Jones), Wiley, **2000**, chapters 3.2.1–3.2.23.
- [11] *Rapid Cycle Real-Time PCR* (Eds.: S. Meuer, C. Wittwer, K. Nakagawara), Springer, Berlin, **2001**.

Detection of Individual p53-Autoantibodies by Using Quenched Peptide-Based Molecular Probes**

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The development of new fast and sensitive assay formats for cancer diagnosis is one of the most challenging tasks in biomedical analysis. Since p53-autoantibodies are independent and highly specific tumor markers with a substantial potential for early-stage diagnosis they have become a focus of modern cancer diagnosis.^[1] Here we report on the development of a sensitive and fast assay for p53-autoantibodies using a novel single-molecule fluorescence spectroscopic method. The method involves detection of reversible fluorescence quenching of dyes by tryptophan residues in labeled short peptides derived from the antibody recognition sequence of the p53 protein. The effectiveness of these peptide-based molecular probes for early-stage diagnosis and follow-up of malignant diseases is demonstrated by the detection of individual p53-autoantibodies directly in sera of cancer patients in the concentration range relevant for clinical tests. Gene mutation in p53—the guardian of the genome^[2]—is the most frequently found abnormality in human cancer^[3] and often triggers the immune system to produce antibodies directed against the nuclear tumor suppressor protein p53—the DNA transcript of the p53 gene. The presence of p53-autoantibodies in human serum validates a malignant disease with a specificity of 100% if autoimmune diseases are neglected.^[1] Commonly, the enzyme-linked immunosorbent assay (ELISA) technique is used as heterogeneous assay format to detect p53-autoantibodies, whose detection limit lies in the range of 10^{−9}–10^{−11}M. The major disadvantages of this method are that it is slow (several hours) and, due to the requirement of recombinant p53, expensive.

Lubin and co-workers have shown that the immune response of cancer patients is not directed against the mutated central segment of the p53 sequence, but rather is directed against a small subset of linear peptide epitopes localized mainly in the N-terminal region (the transactivation domain)

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